

89. Deoxy-nitrosugars

10th Communication¹⁾

Synthesis of Isosteric Phosphonate Analogues of Ulose-1-Phosphates

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A general approach to isosteric phosphonate analogues of ulose-1-phosphates is described. A base-catalysed chain elongation *via* a *Michael* addition of 1-deoxy-1-nitro-sugars **4**, **8**, and **16** to the vinylphosphonate **18** followed by hydrolysis of the nitro adducts gave the analogues of D-ribulose-1-phosphate, D-fructose-1-phosphate, and D-sedoheptulose-1,7-diphosphate **21**, **23**, and **27**, respectively, in high yields.

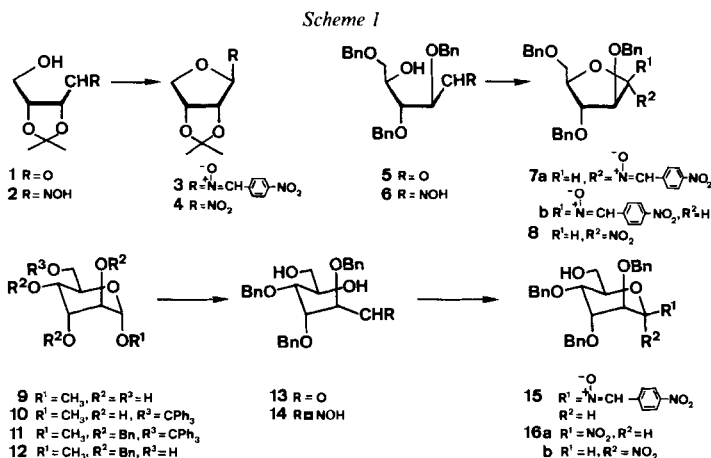
Introduction. – In recent years the interest in analogues of naturally occurring phosphates has focused on isosteric phosphonates; *i.e.* compounds in which the enzymatically cleaved C–O bond of the phosphate monoester is replaced by a non-hydrolysable C–C bond of the analogue [2] [3]. These isosteric phosphonates are potential inhibitors in metabolic processes [2] [4]. Syntheses of sugar phosphonate analogues of reducing phosphates are well-known [2] [5] [6], and syntheses of isosteric phosphonate analogues of aldose-1-phosphates have also been reported [7–10]. A general synthesis of phosphonate analogues of ulose(= ketose)-1-phosphates has so far not been described [4].

The 1-deoxy-1-nitro-aldoses are easily available compounds [11] [12] and potential precursors of isosteric phosphonate analogues of ulose-1-phosphates. Compound **4** is such a 1-deoxy-1-nitrosugar which should easily undergo *Michael* addition to a suitable vinylphosphonate such as **18** (see below, *Scheme 2*). Hydrolysis of the resulting tertiary nitro adducts would lead to the desired phosphonate skeleton such as **19**²⁾. To demonstrate the scope of this approach, we synthesised the phosphonates **21**, **23** [4], and **27** as analogues of D-ribulose-1-phosphate, D-fructose-1-phosphate, and D-sedoheptulose-1,7-diphosphate, respectively. It is well-known that D-ribulose-1,5-diphosphate, D-fructose-1,6-diphosphate, and D-sedoheptulose-1,7-diphosphate play a crucial role in the *Calvin* cycle.

Results. – 2,3-*O*-Isopropylidene-D-erythrose (**1**) [17], prepared from 4,6-*O*-ethylidene-D-glucose [15] [16], was converted to the nitrosugar **4** according to the standard procedure for the preparation of 1-deoxy-1-nitrosugars [12], *i.e.* by transformation *via* the oxime **2** into the nitrone **3** and ozonolysis of this nitrone (*Scheme 1*). The nitro sugar **4** was obtained as a single (α -D) anomer in 56% overall yield from 4,6-*O*-ethylidene-D-glu-

¹⁾ 9th Communication: [1].

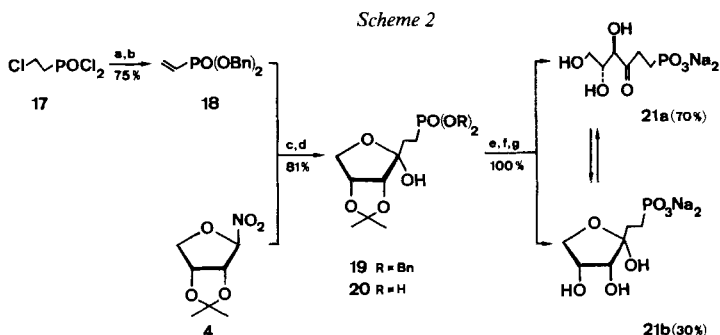
²⁾ The feasibility of this reaction sequence has so far been shown in two cases [13] [14].



cose. Similarly, 2,3,5-tri-*O*-benzyl-D-arabinose (**5**) [18] was converted *via* **6** and **7** to the α -D-configured nitrosugar **8** (76%). Finally, 2,3,4-tri-*O*-benzyl-D-altrose (**13**) was prepared from methyl α -D-altropyranoside (**9**) [19] by tritylation (\rightarrow **10**), benzylation (\rightarrow **11**), detritylation (\rightarrow **12**), and acetylation followed by deacetylation in 45% overall yield. Subsequent transformation into the nitrosugar **16** (82%) again followed the standard procedure (**13** \rightarrow **14** \rightarrow **15** \rightarrow **16**; Scheme 1).

The remaining steps for the preparation of the desired phosphonic acids include a *Michael* addition of the nitrosugars **4**, **8**, and **16**, respectively, to an appropriate vinylphosphonate, the hydrolytic substitution of the NO₂⁻ by an OH-group and a final deprotection. The use of the dibenzyl vinylphosphonate (**18**) as *Michael* acceptor allowed for a facile one-step hydrogenolytic removal of all benzyl groups in the adducts **19**, **22**, and **26**, respectively (see below, Schemes 2-4).

The vinylphosphonate **18** was obtained from (2-chloroethyl)phosphonic dichloride (**17**) [20]³⁾ by esterification with benzyl alcohol and elimination with KOH in EtOH (Scheme 2). In the presence of 0.25 equiv. of Bu₄NF in THF, the nitrosugar **4** reacted



a) BnOH/Et₃N/r.t.; b) KOH/EtOH/10°; c) Bu₄NF/THF/r.t.; d) H₂O/70°; e) H₂/Pd/C/MeOH; f) H₂O/50°; g) Dowex CCR-2 (Bn = CH₂C₆H₅)

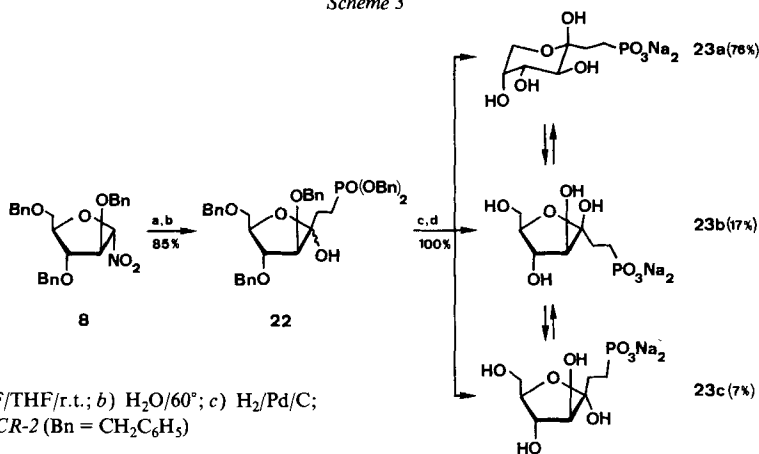
³⁾ We thank Dr. U. Gruntz, Sandoz AG, Basel, for a generous gift of this compound.

rapidly with **18** [14] [21]⁴). Direct hydrolysis of the product at 70° afforded the phosphonate **19** in 81% yield. After hydrogenolysis of **19** (Pd/C, MeOH), the phosphonic acid **20**, which is easily hydrolysable⁵, was converted to the disodium salt **21** by passage through a short column of *Dowex CCR-2* (Na⁺-form) (*Scheme 2*).

In aqueous solution, compound **21** is an equilibrium mixture of the acyclic hydroxyketone **21a** and the cyclic α -D-anomer **21b**. The ¹³C-NMR spectra of **21a** are characterised by a C=O resonance at 214.1 ppm (*d* with ³*J*(C,P) = 14.8 Hz). The corresponding ¹³C-NMR signal of C(3) of **21b** is found at 103.6 ppm, a value typical for an α -D-anomer [22] (*d* with ³*J*(C,P) = 18 Hz). The β -D-anomer of **21** could not be unequivocally identified. The relative amounts of **21a** and **21b** were determined by integration of the signals in the ³¹P-NMR spectrum: **21a** (24.3 ppm)/**21b** (25.3 ppm) = 7:3.

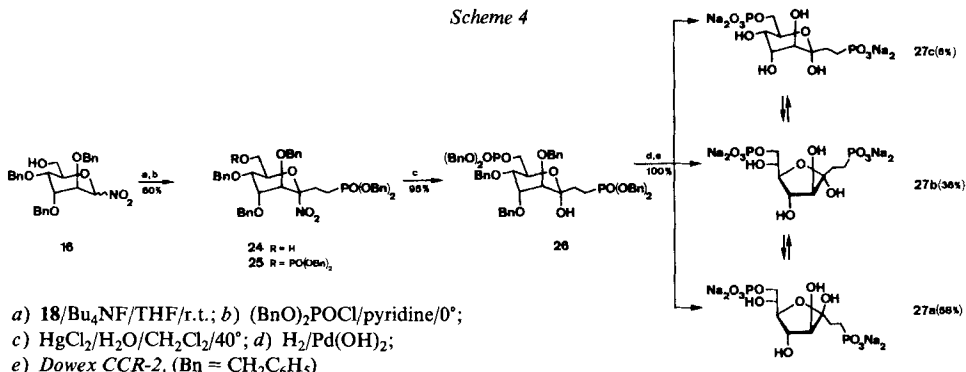
The synthesis of **23**, the analogue of D-fructose-1-phosphate, proceeded in a similar way (*Scheme 3*). *Michael* addition of the tribenzyl-1-deoxy-1-nitroarabinose **8** to **18**, followed by hydrolysis of the addition product gave **22** (83%) as a 38 (α -D): 62 (β -D)

Scheme 3



a) **18**/Bu₄NF/THF/r.t.; b) H₂O/60°; c) H₂/Pd/C;
d) *Dowex CCR-2* (Bn = CH₂C₆H₅)

Scheme 4



a) **18**/Bu₄NF/THF/r.t.; b) (BnO)₂POCl/pyridine/0°;
c) HgCl₂/H₂O/CH₂Cl₂/40°; d) H₂/Pd(OH)₂;
e) *Dowex CCR-2*, (Bn = CH₂C₆H₅)

⁴) No further activation of **18** was necessary. Additions to more highly activated *Michael* acceptors such as ethyl 2-(diethylphosphono)acrylate proceeded in the same way and offered no synthetic advantage.

⁵) Removal of the isopropylidene group prior to debenzylation gave **21** in a very low yield.

mixture of anomers. Hydrogenolysis of **22** ($\text{H}_2/\text{Pd-C}$ in dioxane/ H_2O ⁶⁾) and treatment with *Dowex CCR-2* (Na^+ -form) afforded the sodium salt **23** in nearly quantitative yield. $^{13}\text{C-NMR}$ and $^{31}\text{P-NMR}$ analysis indicated that **23** exists as a 76:17:7 mixture of β -D-pyranose/ β -D-furanose/ α -D-furanose. The $^{13}\text{C-NMR}$ shifts coincide with those determined for D-fructose [22] and D-fructose-1,6-diphosphate [23].

We started the synthesis of the analogue **27** in the usual manner by the addition of the deoxynitroaltrose **16** to the vinylphosphonate **18** (Scheme 4). The attempt to directly hydrolyse the adduct **24** gave mainly the 3,8-anhydro compound. The adduct **24** was, therefore, isolated and phosphorylated with dibenzyl phosphorochloridate [24] to **25** (60% from **16**). The hydrolysis of **25** (wet formamide, MeCN, THF/ H_2O) resulted in decomposition both of the highly labile **25** and of the product **26**. Clean hydrolysis of **25** was, however, observed in the presence of HgCl_2 ($\text{H}_2\text{O}/\text{CH}_2\text{Cl}_2$) to give **26** in 95% yield. Deprotection of **26** was straightforward. Hydrogenolysis with $\text{H}_2/\text{Pd}(\text{OH})_2$ ⁷⁾ [25] at 4 atm in dioxane/ H_2O and conversion of the free acid to the tetrasodium salt afforded the analogue **27** in nearly quantitative yield.

The $^{13}\text{C-NMR}$ data of **27** in D_2O were compared with those reported for sedoheptulose-1,7-diphosphate [26] and found to fit surprisingly well. Our assignments are based on them (see *Exper. Part*). The equilibrium composition of **27** was determined as 58:36:6 for β -D-furanose/ α -D-furanose/ α -D-pyranose by $^{31}\text{P-NMR}$ spectroscopy.

Configuration of the Nitrones and the Nitro Compounds. – Although the preparation of the nitro compounds does not require the isolation of the intermediates, we have isolated the nitrones **3**, **7a**, **7b**, and **15**, respectively, to determine their configuration at the anomeric centre with the help of $^1\text{H-NMR}$ and CD spectroscopy. The nitrone **3** was isolated as a single anomer. The $^1\text{H-NMR}$ spectrum of **3** satisfied *Imbach's* criteria for a β -D(=1,2-*trans*)-configuration, showing a chemical-shift difference for the isopropylidene CH_3 -groups ($\Delta\delta(\text{CH}_3)$) of 0.16 ppm [27] [28] and a *s* for H–C(1) at 5.53 ppm.

Treatment of the arabinose-oxime **6** with *p*-nitrobenzaldehyde gave two compounds which were separated by fractional crystallisation to give the nitrones **7a** and **7b** in a ratio of 93:7. The major anomer shows a *s* at 5.51 ppm in the $^1\text{H-NMR}$ spectrum, whereas the corresponding signal of the minor anomer is found at 5.56 ppm as a *d* with $J(1,2) = 5.4$ Hz. These data confirm the α -D-(=1,2-*trans*)-configuration for the major anomer and the β -D-configuration for the minor anomer.

The nitrone **15** was isolated as a single anomer. The anomeric configuration was, however, not clear from the $^1\text{H-NMR}$ spectrum (axial substituent at C(2)), since the signal for H–C(1) (5.38 ppm, $J(1,2) \leq 0.5$) could correspond to either anomer. A correlation of the CD spectra of the nitrones **3**, **7a**, **7b**, and **15** proved helpful (Table 1). Since the absolute configuration at C(2), C(3), and C(4) is the same in the nitrones **7a**, **7b**, and **15**, respectively, the main contribution to a difference in shape of the CD spectra originates from the configuration at the anomeric centre and the different ring size (furanose and pyranose, resp.). Since the spectra of **7b** and **15** are very similar to each other, the influence of the ring size seems to be very small and the compounds must have the same anomeric configuration. The CD spectrum of the nitrone **3** shows a similar *Cotton* effect to the one of **15** and **7b** in the region of 240–260 nm. The assignment of the β -D-configuration

⁶⁾ Hydrogenation in MeOH or EtOH proceeded much faster, but led to a considerable amount of glycosides, due to the low pH of the solution.

⁷⁾ Hydrogenolysis with 10% Pd/C (*Fluka*) was unsuccessful.

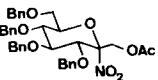
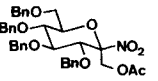
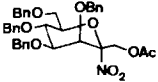
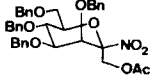
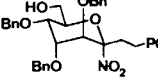
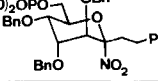
Table 1. UV/CD Spectra (CH_2Cl_2) of Nitrones

Nitrones	3	7a	7b	15
UV, $\lambda_{max}(\epsilon)$	249 (10393)	253 (11511)	253 (11372)	253 (12122)
	348 (17528)	352 (17891)	355 (16774)	352 (18109)
CD, $\lambda_{max}(\Delta\epsilon)$	255 (-0.41)	242 (+1.07)	240 (-1.53)	241 (-1.53)
		272 (-0.26)	284 (-0.09)	276 (+0.51)
	350 (-0.22)	355 (-0.97)	350 (+1.41)	341 (+3.29)

tion to **3** is also consistent with the fact that only the β -D-nitro compound was isolated after ozonolysis.

The configuration of the nitro compounds obtained by ozonolysis of the nitrones, as a rule, correspond to the one of the nitrones, unless the workup included basic conditions (*cf. Exper. Part*). Thus, the nitro- β -D-arabinose obtained from **7** anomerised completely to the more stable α -D-anomer **8** (H-C(1): 5.68 ppm, *s*), and the 1-deoxy-1-nitro- β -D-altriose **16a** anomerised to a 1:2 mixture of **16b** (H-C(1): 5.66 ppm, $J(1,2) = 1.95$) and **16a** (H-C(1): 5.38 ppm, *s*). The α -D-configuration was tentatively assigned to **16b**, based on the chemical shift of H-C(1). *Lemieux* and *Stevens* have shown that the chemical shift of the anomeric H_{eq} is virtually independent of configurational changes at other positions [29], and we consistently found chemical shift values of 5.58–5.77 for nitro α -D-compounds and 5.17–5.38 for the β -D-anomers. This assignment implies that the relative

Table 2. Comparison of Spectral Data of **24** and **25** with those of the Similar Nitro Adducts

Nitro compound	¹³ C-NMR [ppm] anomeric C-atom	IR [cm ⁻¹] $\nu_{as}(NO_2)$	$[\alpha]_D^{25}$	$[\phi]_D^{25}$
	^{a)} 110.62	1552	+55.0°	+352.9°
	^{a)} 109.84	1566	+2.4°	+15.4°
	^{a)} 111.95	1554	+54.5°	+349.7°
	^{a)} 109.46	1577	-38.8°	-249.0°
	24 110.90	1555	+68.6°	+526.6°
	25 110.90	1555	+51.0°	+524.3°

^{a)} See [31].

values of $J(1,2)$ of **16a** and **16b** do not correspond to the mean $J(1,2)$ values indicated by *Altona* and *Haasnoot* [30] for altropyranoses.

The chain elongation of the nitro compounds **4** and **8** by addition to **18** gave in either case mixtures of isomeric nitro adducts, which were directly hydrolysed to **19** and **22**, respectively. *Michael* addition of the mixture of anomers **16a, b** to **18**, however, led to a single isomer **24**, which was isolated. In order to establish its anomeric configuration, we correlated the spectroscopic data of **24** and **25** with the ones of similar nitro adducts prepared from D-glucose and D-mannose, respectively [31] (see *Table 2*), making use of the known fact [32] that in the IR spectrum axial NO₂-groups absorb at lower wave numbers ($\tilde{\nu}_{\text{as}}(\text{NO}_2)$ typically at 1552–1557 cm⁻¹) than equatorial NO₂-groups ($\tilde{\nu}_{\text{as}}(\text{NO}_2)$ typically at 1566–1577 cm⁻¹). The nitro adduct **25** with $\tilde{\nu}(\text{NO}_2)$ 1555 cm⁻¹, therefore, possesses an axial NO₂-group. Moreover, values of the specific rotations of tertiary ethers possessing an axial NO₂-group are more positive than those with an equatorial NO₂-group (*Table 2*). Both criteria indicate the α -D-configuration (axial NO₂-group) for **24** and **25**. The difficult solvolysis of **25** may tentatively be interpreted by invoking a stereoelectronic influence of the vicinal axial (antiperiplanar) benzyloxy group and may be correlated with the difficult oxidation of a D-manno-configured geminal bromo-nitroso compound to the corresponding bromo-nitro compound [32] and the ' Δ -2 effect' [36].

We thank the *Swiss National Science Foundation* and *Sandoz AG*, Basel, for generous support.

Experimental Part

General. See [37]. Compounds on TLC were detected by spraying the plates with a 0.02M soln. of I₂ in 10% aq. H₂SO₄, or by dipping the plates in 10% phosphomolybdic acid in EtOH followed by heating at ca. 200°. Free phosphonic acids on TLC were detected by dipping the plates in vanilline reagent (1 g vanilline, 400 ml MeOH, 100 ml 50% H₂SO₄) and then heated at about 200°. UV spectra were measured with a *Perkin-Elmer-555* spectrophotometer in CH₂Cl₂. IR spectra were measured with a *Perkin-Elmer 298* spectrometer (5% CHCl₃, unless otherwise specified). ¹H-NMR, ¹³C-NMR, and ³¹P-NMR spectra were recorded on a *Varian-HA-100* (¹³C(25.2 MHz)), *Varian-XL-200* (¹H(200 MHz), ¹³C(50 MHz), ³¹P(80 MHz)), or *Bruker-AM-400* spectrometer (¹H(400 MHz), ¹³C(100.6 MHz), ³¹P(160 MHz)); CDCl₃ solns. unless otherwise specified; δ 's in ppm relative to TMS (for ¹H-NMR and ¹³C-NMR) as internal standard or relative to H₃PO₄ (for ³¹P-NMR) as external reference (uncorrected). Mass spectra were recorded on a *Varian 112S* (EI: 70 eV; CI: isobutan) and a *Varian 711* spectrometer (FAB, bombardement with 8-keV Xe-atoms, glycerol matrix).

Dibenzyl Vinylphosphonate (18). At 0°, (2-chloroethyl)phosphonic dichloride (**17**; 10.42 g, 71.38 mmol) was slowly added to benzyl alcohol (17.73 ml, 171.3 mmol) and Et₃N (39.8 ml, 285.5 mmol) in abs. Et₂O (300 ml). After stirring at r.t. for 2 h, H₂O (200 ml) was added and the org. layer was washed with dil. HCl (3 × 100 ml), dil. NaHCO₃ (3 × 100 ml), and H₂O. Drying (MgSO₄) and evaporation *i.v.* afforded 24.0 g of crude dibenzyl (2-chloroethyl)phosphonate. A soln. of this material in abs. EtOH (150 ml) was slowly added to a soln. of KOH (3.21 g, 57.1 mmol) in abs. EtOH (100 ml) at -10°, followed by stirring at 0° for 1 h. Usual workup and chromatography (toluene/AcOEt 3:1) afforded 15.43 g (75%) of **18**. *R_f* (toluene/AcOEt 1:1) 0.36, b.p. 171°/0.23 Torr. IR (film): 3480m (br.), 3090m, 3070m, 3030m, 3010m, 2950m, 2890m, 1960w, 1890w, 1815w, 1720w, 1610m, 1590w, 1500s, 1455s, 1400s, 1380s, 1240s (br.), 1060–960s (br.), 920s, 870–820s (br.), 740s, 700s. ¹H-NMR (200 MHz): 7.33 (m, 10H); 6.60–5.70 (m, 3H); 5.2 (d, $J(\text{H}, \text{P}) = 8.1$, 2 PhCH₂). ¹³C-NMR (25 MHz): 136.0 (d, $J(\text{C}, \text{P}) = 6.4$); 135.5 (dt, $J(\text{C}, \text{P}) = 1.5$); 128.4 (d); 128.2 (d); 127.7 (d); 125.5 (dd, $J(\text{C}, \text{P}) = 185.2$); 67.3 (dt, $J(\text{C}, \text{P}) = 5.6$). ³¹P-NMR (80 MHz): 18.9. Anal. calc. for C₁₆H₁₇O₃P (288.28): C 66.66, H 5.94, P 10.74; found: C 66.35, H 5.83, P 10.70.

⁸⁾ A similar difference for $\tilde{\nu}(-\text{N}=\text{C})$ in the IR spectra of anomeric glycosyl isocyanides has been reported [33–35].

2,3-O-Isopropylidene-D-erythrose (1). A mixture of crude D-erythrose (15.09 g, 0.1 mol) [16], abs. acetone (400 ml), and anh. FeCl₃ (1 g, 5 mol-%) [17] was stirred at r.t. under anh. conditions for 48 h. Then Na₂CO₃ · 10H₂O (50 g) was added, and stirring was continued for 2 h at r.t. After filtration through *Celite* and drying (MgSO₄) the soln. was concentrated *i.v.* to afford 18.2 g of crude **1** [38] as an oil. *R_f* (hexane/AcOEt 1:2) 0.46.

2,3-O-Isopropylidene-D-erythrose-oxime (2). A soln. of crude **1** (18.2 g) in abs. MeOH (50 ml) was added to a mixture of NH₂OH · HCl (16.07 g, 231 mmol) and NaOCH₃ (11.24 g, 208 mmol) in abs. MeOH (250 ml). After stirring at r.t. for 2 h, MeOH was removed *i.v.* (bath temp. 40°). The residue was diluted with H₂O and AcOEt, and the H₂O-phase was extracted with AcOEt (4 × 100 ml). Drying (MgSO₄) and concentration *i.v.* afforded 20.83 g of crude **2** as a thick sirup. For analysis, a small amount of **2** was crystallised (hexane/AcOEt). M.p. 90–94°, *R_f* (hexane/AcOEt 1:2) 0.23. IR (KBr): 3400s, 3210s, 3090s, 3015m, 3000s, 2940s, 2900m, 1660w, 1630w, 1440m, 1420m, 1385s, 1340m, 1300m, 1270s, 1260s, 1230s, 1165s, 1130m, 1110m, 1080s, 1050s, 990m, 960m, 930m, 915s, 890s, 840s, 800m, 705m. ¹H-NMR (200 MHz): (Z)-oxime: 6.95 (*d*, *J* = 4.53, H–C(1)); 5.29 (*dd*, *J* = 7.1, 4.5, H–C(2)); 4.56–4.48 (*m*, H–C(3)); 3.71–3.50 (*m*, 2H–C(4)); 2.8 (br. *s*, OH); 1.53, 1.39 (2*s*, each 3H, 2 CH₃); (E)-oxime: 7.46 (*d*, *J* = 7.46, H–C(1)); 4.74 (*dd*, *J* = 7.13, H–C(2)); 4.42–4.34 (*m*, H–C(3)); 3.71–3.50 (*m*, 2H–C(4)); 2.80 (br. *s*, OH); 1.53, 1.39 (*s*, each 3H, 2 CH₃); integral over H–C(1) signals: (E/Z) = 1:2. ¹³C-NMR (25 MHz): (Z)-oxime: 150.4 (*d*); 110.2 (*s*); 79.6 (*d*); 72.5 (*d*); 62.4 (*t*); 27.6, 25.1 (2*q*); (E)-oxime: 148.5 (*d*); 110.5 (*s*); 79.8 (*d*); 76.1 (*d*); 61.5 (*t*); 27.9, 25.4 (2*q*). MS (EI): 160 (5, *M*⁺ – 15), 98 (9), 70 (21), 59 (65), 43 (100). Anal. calc. for C₇H₁₃NO₄ (175.19): C 47.99, H 7.48, N 7.99; found: C 47.78, H 7.50, N 7.87.

N-(p-Nitrophenylmethylidene)-2,3-O-isopropylidene-β-D-erythrosufuranosylamine N-Oxide (3). A mixture of crude **2** (20.83 g), *p*-nitrobenzaldehyde (26.2 g, 173 mmol), TsOH (50 mg) and Drierite (50 g) in abs. CH₂Cl₂ (250 ml) was stirred at r.t. under anh. conditions for 2 h and then filtered through *Celite*. For analysis, a small amount of **3** was washed with 10% aq. NaHSO₃ soln. (4×) and H₂O (3×) and then dried (MgSO₄). Concentration *i.v.* and crystallisation (Et₂O/hexane) afforded analytically pure **3**. M.p. 155–157°, *R_f* (hexane/AcOEt 1:2) 0.52, [α]_D²⁵ = –133.8° (CHCl₃, *c* = 0.93). UV: 249 (10393), 348 (17528). IR (KBr): 3100w, 3090w, 3000m, 1600s, 1565s, 1515s, 1415m, 1385s, 1355s, 1220s, 1140s, 1110s, 1100s, 1055s, 865s, 755m, 730m, 695m. ¹H-NMR (200 MHz): 8.41, 8.26 (2*d*, *AA'**BB'*, *J* = 9, 2H each, C₆H₄); 7.78 (*s*, NO=CH); 5.53 (*s*, H–C(1)); 5.30 (*d*, *J* = 5.8, H–C(2)); 5.00 (*dd*, *J* = 5.8, 3.4, H–C(3)); 4.46 (*dd*, *J* = 10.2, 3.8, H_{exo}–C(4)); 4.35 (*d*, *J* = 10.2, H_{endo}–C(4)); 1.54, 1.38 (2*s*, each 3H, 2 CH₃). ¹³C-NMR (50 MHz): 147.8 (*s*); 135.1 (*s*); 130.0 (*d*); 129.0 (*d*); 123.6 (*d*); 113.1 (*s*); 104.4 (*d*); 84.4 (*d*); 80.1 (*d*); 76.8 (*t*); 26.2 (*q*); 24.6 (*q*). MS (CI): 309 (*M*⁺ + 1). Anal. calc. for C₁₄H₁₆N₂O₆ (308.29): C 54.54, H 5.23, N 9.08; found: C 54.45, H 5.17, N 9.20.

1-Deoxy-2,3-O-isopropylidene-1-nitro-β-D-erythrose (4). The filtered CH₂Cl₂ soln. of the crude nitrone **3** was diluted with CH₂Cl₂ (250 ml) and cooled to –50°. O₃ was bubbled through the soln. until disappearance of **3** (monitored by TLC (hexane/AcOEt 1:2)). The soln. was purged with N₂ (30 min at –50°, then 30 min at r.t.), washed with 10% aq. NaHSO₃ (3 × 200 ml) and H₂O (3 × 200 ml), dried (MgSO₄), and concentrated *i.v.* Chromatography (hexane/AcOEt 10:1) and crystallisation (Et₂O/hexane) afforded 11.12 g (52.6%, from 4,6-O-ethylidene-D-glucose) of **4**. M.p. 42.5–43.5°, *R_f* (hexane/AcOEt 10:1) 0.18, [α]_D²⁵ = –116.0° (*c* = 0.96, CHCl₃). IR: 3015m, 3000s, 2860s, 2840s, 2800m, 1775s, 1460s, 1385s, 1380s, 1350m, 1310m, 1270s, 1150s, 1105s, 1080s, 1050s, 980s, 940s, 860s. ¹H-NMR (200 MHz): 5.73 (*s*, H–C(1)); 5.02 (*d*, *J* = 6.0, H–C(2)); 4.93 (*dd*, *J* = 6.0, 3.0, H–C(3)); 4.48 (*dd*, *J* = 11.0, 3.0, H_{exo}–C(4)); 4.40 (*d*, *J* = 11.0, H_{endo}–C(4)); 1.52 (*s*, CH₃); 1.36 (*s*, CH₃). ¹³C-NMR (25.2 MHz): 113.9 (*s*); 111.5 (*d*); 85.9 (*d*); 79.4 (*d*); 78.2 (*t*); 26.2 (*q*); 24.7 (*q*). MS (CI): 190 (*M*⁺ + 1), 174 (*M*⁺ – 15), 143 (*M*⁺ – 46). Anal. calc. for C₇H₁₁NO₅ (189.2): C 44.45, H 5.86, N 7.40; found: 44.69, H 6.00, N 7.68.

Dibenzyl 1,2-Dideoxy-4,5-O-isopropylidene-α-D-erythro-3-hexulofuranose-1-phosphonate (19). Bu₄NF · 3H₂O (2.08 g, 6.60 mmol) was added in one portion to a stirred mixture of **4** (5.00 g, 26.43 mmol) and **18** (7.62 g, 26.43 mmol) in THF (80 ml) at 0°. After having stirred at r.t. for 1 h, H₂O (25 ml) and sat. aq. NaHCO₃ soln. (20 ml) were added and the mixture was heated at 70° for 2–3 h. Usual workup, chromatography (hexane/AcOEt 1:2) and crystallisation (Et₂O/hexane) afforded 9.60 g (81%) of **19**. M.p. 82.5–83.5°, *R_f* (hexane/AcOEt 1:2) 0.16, [α]_D²⁵ = –25.1° (*c* = 1.1, CHCl₃, *t* = 45 min). IR: 3300s (br.), 3090w, 3070w, 3030w, 2990s, 2940s, 2880m, 1950w, 1880w, 1810w, 1605w, 1590w, 1495m, 1455s, 1380s, 1375s, 1320m, 1270–1190s (br.), 1095s, 1060–970s (br.), 920s, 860s. ¹H-NMR (400 MHz): α-D-anomer: 7.35 (*m*, 10H); 5.08–4.92 (*m*, 2 PhCH₂); 4.84 (*dd*, *J* = 6.0, 3.9, H–C(5)); 4.38 (*d*, *J* = 6.0, H–C(4)); 4.06 (*dd*, *J* = 10.3, 3.9, H_{exo}–C(6)); 3.90 (*d*, *J* = 10.3, H_{endo}–C(6)); 2.34–1.88 (*m*, 2H–C(1), 2H–C(2)); 1.44 (*s*, CH₃); 1.31 (*s*, CH₃); β-D-anomer: 4.68 (*m*, H–C(5)); 4.61 (*d*, *J* = 4.6, H–C(4)); 3.84 (*d*, *J* = 12, H_{endo}–C(6)); 3.46 (*dd*, *J* = 12, 4.7, H_{exo}–C(6)); 1.75 (*m*, 2H–C(2), 2H–C(1)); 1.54, 1.35 (2*s*, each 3H, 2 CH₃). ¹³C-NMR (25.2 MHz): α-D-anomer: 136.1 (*d*, *J*(C, P) = 6.3); 128.4 (*d*); 128.2 (*d*); 127.7 (*d*); 112.1 (*s*); 106.0 (*d*, *J*(C, P) = 14.3); 84.6 (*d*); 80.8 (*d*); 70.8 (*t*); 67.4 (*dt*, *J*(C, P) = 6.4); 67.3 (*dt*, *J*(C, P) = 6.4); 27.9 (*dt*, *J*(C,

P) = 4.7, 26.3 (q); 24.9 (q); 20.4 (dt, J(C, P) = 141.5); β -D-anomer: 128.6 (d); 128.0 (d); 127.9 (d); 113.5 (s); 81.1 (d); 79.8 (d); 68.2 (t); 28.1 (dt, J(C, P) = 2.2); 26.1 (q); 19.6 (dt, J(C, P) = 143.3). $^{31}\text{P-NMR}$ (80 MHz): 35.6 (s), 34.3 (m), $\alpha/\beta = 91.2:8.8$. MS (CI): 449 ($M^+ + 1$), 431 ($(M^+ + 1) - 18$), 373, 341. Anal. calc. for $\text{C}_{23}\text{H}_{29}\text{O}_7\text{P}$ (448.46): C 61.60, H 6.52, P 2.10; found: C 61.64, H 6.42, P 1.95.

1,2-Dideoxy-4,5-O-isopropylidene- α -D-erythro-3-hexulofuranose-1-phosphonic Acid (20). Ester **19** (1.00 g, 2.23 mmol) was treated with H_2 and 10% Pd/C (100 mg, Fluka) in MeOH (20 ml) at r.t. and under normal pressure for 2 h. Filtration and evaporation afforded 589 mg (100%) of crystalline **20**. M.p. 128–129°, R_f (PrOH/NH₃/H₂O 4:3:1) 0.54, $[\alpha]_{\text{D}}^{25} = -54.4^\circ$ ($c = 0.88$, MeOH, $t = 13$ min). IR (KBr): 3380s (br.), 2995s, 2920s, 2700s (br.), 2300m (br.), 1630w (br.), 1430m, 1390m, 1380m, 1325m, 1280s, 1240s, 1210s, 1180s, 1150s, 1095s, 1050s, 1000s, 940s, 890m, 860s, 835m, 820m, 785m. $^1\text{H-NMR}$ (200 MHz, (D₆)DMSO): 7.80 (br. s, 3 OH); 4.80 (dd, $J = 5.9, 3.7$, H-C(5)); 4.23 (d, $J = 5.9$, H-C(4)); 3.82 (dd, $J = 10.0, 3.7$, H_{exo}-C(6)); 3.70 (d, $J = 10.0$, H_{endo}-C(6)); 1.95–1.50 (m, 2H-C(1), 2H-C(2)); 1.35 (s, CH₃); 1.24 (s, CH₃). $^{13}\text{C-NMR}$ (100 MHz, (D₆)DMSO): 111.1 (s); 105.8 (d, J(C, P) = 15.6); 84.5 (d); 80.2 (d); 69.9 (t); 28.4 (t); 26.2 (q); 24.8 (q); 22.0 (dt, J(C, P) = 145.8). $^{31}\text{P-NMR}$ (160 MHz, (D₆)DMSO): 29.5. MS (FAB): 291 ($M + \text{Na}$), 313 ($M - 1 + 2\text{Na}$), 335 ($M - 2 + 3\text{Na}$). Anal. calc. for $\text{C}_9\text{H}_{17}\text{O}_7\text{P}$ (268.21): C 40.31, H 6.39, P 11.55; found: C 39.95, H 6.35, P 11.30.

Sodium 1,2-Dideoxy-D-erythro-3-hexulose-1-phosphonate (21). A soln. of **20** (3.00 g, 11.2 mmol) in 200 ml H₂O was kept at 50° for 24 h. The mixture was concentrated *i.v.* to 40 ml and passed through a column of Dowex CCR-2 (Na⁺-form). The effluent was lyophilised and then dried *i.v.* over P₂O₅. Yield: 3.10 g (100%) of **21**. R_f (PrOH/NH₃/H₂O 4:3:1) 0.35, $[\alpha]_{\text{D}}^{25} = -16.3^\circ$ ($c = 1.19$, H₂O). $^1\text{H-NMR}$ (200 MHz, D₂O): 4.38 (d, $J = 5.3$, H-C(4)); 4.05 (q, $J = 5.3$, H-C(5)); 3.66 (m, 2H-C(6)); 2.87 (m, 1H); 1.76 (m, 3H), other isomers: 4.5–3.5 (m). $^{13}\text{C-NMR}$ (100 MHz, D₂O): acyclic form: 214.1 (d, J(C, P) = 14.8, C(3)); 77.6 (d); 72.7 (d); 61.6 (t, C(6)); 34.8 (t, C(2)); 22.2 (dt, J(C, P) = 135.2, C(1)); α -D-anomer: 103.6 (d, J(C, P) = 18.0, C(3)); 77.2 (d); 71.9 (d); 62.6 (t, C(6)); 33.8 (t, C(2)); 22.5 (dt, J(C, P) = 133.0, C(1)). $^{31}\text{P-NMR}$ (160 MHz, D₂O): 25.3 (α -D-anomer); 24.3 (acyclic form); integral approx. 3:7 for α -D/acyclic form. MS (FAB): 387 ($M + \text{Na} + \text{gly}$), 365 ($M + 1 + \text{gly}$), 343 ($M + 2 - \text{Na} + \text{gly}$), 321 ($M + 3 - 2\text{Na} + \text{gly}$), 295 ($M + \text{Na}$), 273 ($M + 1$), 251 ($M + 2 - \text{Na}$), 229 ($M + 3 - 2\text{Na}$). Anal. calc. for $\text{C}_6\text{H}_{11}\text{Na}_2\text{O}_7\text{P}$ (272.10): C 26.48, H 4.07, P 11.38; found: C 26.57, H 4.30, P 11.20.

2,3,5-Tri-O-benzyl-D-arabinose-oxime (6). In one portion, 2,3,5-tri-O-benzyl-D-arabinose (5; 2.00 g, 47.56 mmol) [18] was added to a soln. of NaOMe (from 1.64 g (71.34 mmol) of Na) and H₂NOH·HCl (6.61 g, 95.12 mmol) in abs. MeOH (130 ml). The heterogenous mixture was then stirred at r.t. for 2 h under anh. conditions. Evaporation of MeOH *i.v.*, extraction of the residue with CH₂Cl₂ (3 × 200 ml), drying (MgSO₄), and evaporation afforded 22 g (quantitative) of **6** as a viscous sirup. R_f (toluene/AcOEt 2:1) 0.31. For analysis, a small amount of **6** was purified by chromatography (toluene/AcOEt 2:1). R_f (toluene/AcOEt 2:1) 0.31, $[\alpha]_{\text{D}}^{25} = -8.67^\circ$ ($c = 2.41$, CH₂Cl₂). IR: 3580s, 3350s (br.), 3080m, 3060m, 3020m, 3000s, 2910m, 2860m, 1950w, 1870w, 1810w, 1605w, 1580w, 1490w, 1450s, 1390m, 1250–1200m (br.), 1080s (br.), 1025s, 910m. $^1\text{H-NMR}$ (200 MHz): (*E*)-oxime: 8.48 (s, =NOH); 7.48 (d, $J = 7.92$, H-C(1)); 7.37–7.22 (m, 15H); 4.50 (m, 3 AB, 3 PhCH₂); 4.31 (dd, $J = 3.5, 8$, H-C(2)); 4.06 (m, H-C(4)); 3.66 (m, 3H, H-C(3), 2H-C(5)); 3.15 (d, $J = 6$, OH-C(4)); (*Z*)-oxime: 8.68 (s, =NOH); 7.37–7.22 (m, 15H); 7.01 (d, $J = 6.1$, H-C(1)); 5.90 (dd, $J = 6.1, 3.0$, H-C(2)); 4.50 (m, 3 AB, 3 PhCH₂); 4.06 (m, H-C(4)); 3.88 (dd, $J = 7.5, 3.0$, H-C(3)); 3.66 (m, 2H-C(5)); 2.85 (d, $J = 6.0$, OH-C(4)). $^{13}\text{C-NMR}$ (50 MHz): (*E*)-oxime: 149.9 (s); 137.7–137.3 (3s); 128.4–127.7 (9d); 80.2 (d); 76.7 (d); 74.2 (t); 73.5 (t); 71.4 (t); 70.9 (t); 69.9 (d); (*Z*)-oxime: 152.1 (s); 137.7–137.3 (3s); 128.4–127.7 (9d); 79.2 (d); 73.4 (d); 72.5 (t); 71.9 (d); 71.0 (t); 69.8 (d); isomer ratio (*E*)/(*Z*) = 2:1. MS (EI): 435 (M^+), 418 ($M^+ - 17$), 344 ($M^+ - 91$). Anal. calc. for $\text{C}_{26}\text{H}_{29}\text{NO}_5$ (435.52): C 71.70, H 6.71, N 3.22; found: C 71.46, H 6.93, N 3.10.

N-(p-Nitrophenylmethylidene)-2,3,5-tri-O-benzyl-D-arabinofuranosylamine N-Oxide (7). A mixture of the crude **6** (22 g), *p*-nitrobenzaldehyde (8.62 g, 57.07 mmol), TsOH (100 mg) and Drierite (20 g) in abs. CH₂Cl₂ (200 ml) was stirred at r.t. for 2 h. Drierite was removed by filtration through Celite and the filtrate was concentrated *i.v.* For analysis, a small amount of **7** was washed with 10% aq. NaHSO₃ soln. (4×) and H₂O (3×), and then dried (MgSO₄). Concentration *i.v.* and fractional crystallisation (Et₂O/hexane) afforded the α -D-nitronone **7a** as the main component and a small amount of the β -D-nitronone **7b** ($\alpha/\beta = 93:7$).

α -D-Nitronone 7a. M.p. 101–103°, R_f (toluene/AcOEt 2:1) 0.61, $[\alpha]_{\text{D}}^{25} = -1.4^\circ$ ($c = 0.95$, CHCl₃). UV: 253 (11511), 352 (17891). IR (KBr): 3100w, 3070w, 3030w, 2960w, 2880m, 1600m, 1570m, 1520s, 1500m, 1470m, 1455m, 1415m, 1350s, 1205m, 1135m, 1100s, 1000m, 880m, 865m, 840m, 760s, 740s, 705s. $^1\text{H-NMR}$ (200 MHz): 8.41, 8.25 (d, $J = 9$, AA'BB', C₆H₄); 7.92 (s, NO=CH); 7.44–7.02 (m, 15H); 5.51 (s, H-C(1)); 4.85, 4.69 (AB, PhCH₂); 4.70 (m, 1H); 4.63 (m, 1H); 4.64, 4.55 (AB, PhCH₂); 4.42, 4.34 (AB, PhCH₂); 4.01 (dd, $J = 2.9, 1.5$, H-C(3)); 3.68 (m, 2H-C(5)). $^{13}\text{C-NMR}$ (50 MHz): 147.8 (s); 137.6 (s); 137.0 (s); 136.8 (s); 135.4 (s); 129.1–127.4 (12d); 123.5 (d); 102.5 (d); 86.3 (d); 85.9 (d); 82.4 (d); 73.4 (t); 72.2 (t); 71.6 (t); 69.6 (t). MS (CI): 369 ($M^+ + 1$). Anal. calc. for $\text{C}_{33}\text{H}_{32}\text{N}_2\text{O}_7$ (568.63): C 69.70, H 5.67, N 4.93; found: C 69.53, H 5.70, N 4.87.

β -D-Nitron 7b. M.p. 74–77°, R_f (toluene/AcOEt 2:1) 0.49, $[\alpha]_D^{25} = +27.7^\circ$ ($c = 0.53$, CHCl_3). UV: 253 (11371), 355 (16774). IR (KBr): 3080w, 3040w, 2905m, 2890m, 1600m, 1575m, 1520s, 1455m, 1340s, 1260m, 1220m, 1140s, 1100s, 1035m, 870m, 760m, 750m, 705m, 700m. $^1\text{H-NMR}$ (200 MHz): 8.09 (s, $AA'A''$, C_6H_4); 7.93 (s, $\text{NO}=\text{CH}$), 7.37–7.24 (m, 15H); 5.56 (d, $J = 5.4$, H–C(1)); 4.78, 4.57 (AB, PhCH_2); 4.64, 4.53 (AB, PhCH_2); 4.53 (s, PhCH_2); 4.46 (m, 1H); 4.23 (m, 2H); 3.78 (m, 2H). $^{13}\text{C-NMR}$ (50 MHz): 147.5 (s); 137.5, 137.4, 137.1 (3s); 135.6 (s); 130.2 (d); 129.1 (d); 128.4–127.6 (7d); 123.4 (d); 98.6 (d); 82.8, 82.0, 79.9 (3d); 74.0, 73.4, 72.3 (3t); 68.8 (t). MS (CI): 569 ($M^+ + 1$). Anal. calc. for $\text{C}_{33}\text{H}_{32}\text{N}_2\text{O}_7$ (568.63): C 69.70, H 5.67, N 4.93; found: C 69.75, H 5.72, N 4.95.

2,3,5-Tri-O-benzyl-1-deoxy-1-nitro- α -D-arabinofuranose (8). A soln. of crude **7** in CH_2Cl_2 (350 ml) was treated with O_3 at -78° (the reaction was monitored by TLC and immediately stopped when **7** had disappeared). The soln. was purged with N_2 (first at -78° for 30 min, then at r.t. for 30 min), washed with 10% aq. NaHSO_3 (3×200 ml) and H_2O (3×200 ml). Drying (MgSO_4) and concentration *i.v.* afforded a sirup, which was treated with NaBH_4 (899 mg, 23.78 mmol) and anh. LiCl (1.00 g, 23.78 mmol) in abs. diglyme (80 ml) at r.t. for 3 h⁹). Usual workup and chromatography (toluene/AcOEt 70:1) afforded 16.25 g (76%) of **8** as an oil. R_f (toluene/AcOEt 30:1) 0.41, $[\alpha]_D^{20} = +69.5^\circ$ ($c = 0.68$, CHCl_3). IR: 3090m, 3060m, 3030m, 3010m, 2920m, 2870m, 1950w, 1875w, 1810w, 1605w, 1565s, 1495s, 1450s, 1365s, 1310m, 1250m, 1160s, 1110s, 1030s, 910m, 690s. $^1\text{H-NMR}$ (200 MHz): 7.35–7.18 (m, 15H); 5.68 (s, H–C(1)); 4.72 (d, AB, $J = 12.0$, 1H, PhCH_2); 4.65 (q, $J = 5.2$, H–C(4)); 4.57 (d, AB, $J = 12.0$, 1H, PhCH_2); 4.55 (s, PhCH_2); 4.41 (br. s, and d, $J = 3.0$, PhCH_2 , H–C(2), resp.); 4.03 (dd, $J = 5.2$, 2.5, H–C(3)); 3.64 (d, $J = 5.2$, 2H–C(5)). $^{13}\text{C-NMR}$ (25.2 MHz): 137.6 (s); 136.8 (s); 136.2 (s); 128.9–127.6 (8d); 108.9 (d); 87.8 (d); 85.7 (d); 82.1 (d); 73.4 (t); 72.4 (t); 72.0 (t); 68.7 (t). MS (CI): 449 (M^+), 402 ($M^+ - \text{NO}_2$), 358 ($M^+ - 91$). Anal. calc. for $\text{C}_{26}\text{H}_{27}\text{NO}_6$ (449.51): C 69.47, H 6.05, N 3.12; found: C 69.23, H 6.08, N 3.33.

Dibenzyl 4,5,7-Tri-O-benzyl-1,2-dideoxy-D-arabino-3-heptulofuranose-1-phosphonat (22). At 0° , $\text{Bu}_4\text{NF} \cdot 3\text{H}_2\text{O}$ (1.42 g, 4.52 mmol) was added in one portion to **8** (8.13 g, 18.09 mmol) and **18** (5.47 g, 18.99 mmol) in dry THF (80 ml). After stirring at r.t. for 1 h, H_2O (80 ml) and sat. NaHCO_3 (20 ml) was added and the mixture heated at 60° for 24 h. Usual workup and chromatography (toluene/AcOEt 2:1) afforded 10.67 g (85%) of **22** as a colorless oil. R_f (toluene/AcOEt 2:1) 0.11, $[\alpha]_D^{25} = +7.2^\circ$ ($c = 0.54$, CHCl_3). IR: 3600–3200m (br.), 3090m, 3070m, 3030m, 2990m, 2920m, 2860m, 1955w, 1875w, 1810w, 1720w, 1605w, 1580w, 1495m, 1450s, 1360m, 1250–1200s (br.), 1100–1000s (br.), 915m, 855m. $^1\text{H-NMR}$ (400 MHz): α -D-isomer: 7.40–7.10 (m, 25H); 5.05–4.91 (m, ($\text{PhCH}_2\text{O}_2\text{P}$)); 4.62–4.32 (m, 3 AB, 3 PhCH_2); 4.22 or 4.13 (s, OH); 4.12 (t, $J = 4.5$, H–C(5)); 3.98 (q, $J = 4.4$, H–C(6)); 3.76 (d, $J = 4.5$, H–C(4)); 3.54 (dd, $J = 10.2$, 4.5, H–C(7)); 3.47 (dd, $J = 10.2$, 4.0, H–C(7)); 2.20–1.90 (m, 2H–C(2), 2H–C(1)); β -D-isomer: 7.40–7.10 (m, 25H); 5.05–4.91 (m, ($\text{PhCH}_2\text{O}_2\text{P}$)); 4.62–4.32 (m, 3 AB, 3 PhCH_2); 4.35 (m, H–C(6)); 4.22 or 4.13 (s, OH); 3.92 (dd, $J = 2.7$, 1.6, H–C(5)); 3.78 (d, $J = 1.6$, H–C(4)); 3.55 (d, $J = 9.8$, H–C(7)); 3.43 (dd, $J = 9.8$, 7.0, H–C(7)); 2.20–1.90 (m, 2H–C(2), 2H–C(1)). $^{13}\text{C-NMR}$ (100 MHz): 138.1–136.4 (7s), 128.5–127.6 (13d); α -D-isomer: 106.5 (d, $J(\text{C}, \text{P}) = 16.2$); 85.9 (d); 82.1 (d); 81.5 (d); 73.3 (t); 71.8 (t); 71.7 (t); 70.2 (t); 67.2 (dt, $J(\text{C}, \text{P}) = 6.5$); 27.7 (dt, $J(\text{C}, \text{P}) = 3.6$); 20.3 (dt, $J(\text{C}, \text{P}) = 141.9$); β -D-isomer: 102.7 (d, $J(\text{C}, \text{P}) = 17.7$); 86.2 (d); 83.8 (d); 80.1 (d); 73.5 (t); 72.5 (t); 72.1 (t); 70.8 (t); 67.1 (dt, $J(\text{C}, \text{P}) = 6.5$); 31.2 (dt, $J(\text{C}, \text{P}) = 3.9$); 19.8 (dt, $J(\text{C}, \text{P}) = 142.8$). $^{31}\text{P-NMR}$ (160 MHz): 35.1 (α); 34.5 (β), integral $\alpha/\beta = 38:62$. Anal. calc. for $\text{C}_{42}\text{H}_{45}\text{O}_8\text{P}$ (708.80): C 71.17, H 6.40, P 4.37; found: C 70.48, H 6.66, P 4.20.

Disodium 1,2-Dideoxy-D-arabino-3-heptulose-1-phosphonate (23). A soln. of **22** (4.31 g, 6.08 mmol) in 100 ml of dioxane/ H_2O 1:1 was treated with 10% Pd/C (1 g, Fluka) under H_2 at normal pressure. After 3 h, the catalyst was removed by filtration and the soln. was concentrated to half of the volume. After dilution with H_2O (50 ml) and addition of fresh catalyst (1 g), the hydrogenation was continued for 3 h. Filtration, concentration *i.v.* to 40 ml, treatment with Dowex CCR-2 (Na^+ -form) and lyophilisation afforded 1.87 g (100%) of **23**. R_f ($\text{PrOH}/\text{NH}_3/\text{H}_2\text{O}$ 4:3:1) 0.24, $[\alpha]_D^{25} = -38.1^\circ$ ($c = 0.86$, H_2O). $^1\text{H-NMR}$ (200 MHz, D_2O): 4.06–3.63 (m, 5H); 1.95 (m, 2H); 1.59 (m, 2H). $^{13}\text{C-NMR}$ (50 MHz, D_2O): due to line broadening of ^{13}C -signals of **23** (pH 8, D_2O), the data of the free acid are reported (pH 1–2, D_2O): β -D-pyranose: 98.9 (d, $J(\text{C}, \text{P}) = 17.4$, C(3)); 70.5 (d, C(5)); 70.1 (d, C(6)); 69.5 (d, C(4)); 63.9 (t, C(7)); 31.3 (t, C(2)); 20.8 (dt, $J(\text{C}, \text{P}) = 136.2$, C(1)); β -D-furanose: 102.4 (d, $J(\text{C}, \text{P}) = 19.0$, C(3)); 80.9 (d, C(6)); 78.9 (d, C(4)); 75.2 (d, C(5)); 62.9 (t, C(7)); 31.3 (t, C(2)); 21.2 (dt, $J(\text{C}, \text{P}) = 136.8$, C(1)); α -D-furanose: 106.3 (d, $J(\text{C}, \text{P}) = 19.7$, C(3)); 82.6 (d, C(4)); 82.1 (d, C(6)); 77.5 (d, C(5)); 61.8 (t, C(7)); C(2) and C(1) not visible; α -D-pyranose: 99.6 (d, $J(\text{C}, \text{P}) = 17.3$, C(3)); additional signals at 77.0 (d), 72.5 (d), 71.6 (d); 71.1 (d); 66.7 (d); 31.6 (t); and 31.0 (t). $^{31}\text{P-NMR}$ (160 MHz, D_2O , integral): 23.1 (**23a**, 76%); 22.6 (**23b**, 17%); 21.5 (**23c**, 7%). MS (FAB): 303 ($M^+ + 1$). Anal. calc. for $\text{C}_7\text{H}_{13}\text{Na}_2\text{O}_8\text{P}$ (302.13): C 27.82, H 4.33, P 10.25; found: C 27.30, H 4.45, P 9.86.

Methyl 6-O-Triyl- α -D-altropyranoside (10). A mixture of methyl α -D-altropyranoside **9** [19] (20.68 g, 106.5 mmol), trityl chloride (44.53 g, 159.7 mmol), and 4-(N,N-dimethylamino)pyridine (650 mg, 5 mol-%) in abs.

⁹) To transform any benzoates formed by overoxidation into the corresponding easily removed alcohols [12].

pyridine (50 ml) and abs. CHCl_3 (100 ml) was stirred at r.t. for 12 h. Dilution with H_2O (100 ml), extraction with CHCl_3 (4×150 ml), drying (MgSO_4), and evaporation *i.v.*, followed by co-evaporation with EtOH and toluene ($5 \times$) afforded a yellow sirup. Chromatography (hexane/AcOEt 1:6) and drying *i.v.* gave 36.6 g (79%) of **10** as a slightly yellow foam. R_f (hexane/AcOEt 1:6) 0.3.

Methyl 2,3,4-Tri-O-benzyl- α -D-altropyranoside (12). NaH (4.08 g, 169.89 mmol; Fluka, 60% NaH in oil) was washed with dry hexane under N_2 and then added in one portion to a soln. of **10** (14.54 g, 33.31 mmol) in abs. DMF (200 ml). After stirring at r.t. for 30 min, benzyl bromide (29.7 ml, 249.82 mmol) was added dropwise over 30 min to the cooled (0°) suspension, followed by stirring at r.t. for 24 h. Excess NaH was then destroyed with abs. MeOH (15 ml) and the mixture treated with 13 g of thiourea at r.t. for 2 h. Evaporation of DMF *i.v.* and usual workup furnished 31.1 g of crude **11**. This material was treated with ZnBr_2 (150 g, 0.66 mol) [39] in abs. CH_2Cl_2 (70 ml) for 3 h under vigorous stirring. Cautious addition of abs. MeOH (30 ml) followed by H_2O (100 ml), usual workup, and chromatography (toluene/AcOEt 3:1) afforded 13.30 g (86%) of **12** as an oil. R_f (toluene/AcOEt 3:1) 0.14, $[\alpha]_D^{25} = +82.2^\circ$ ($c = 1.03$, CHCl_3). IR: 3590m, 3510m (br.), 3090m, 3070m, 3030m, 3000s, 2930s, 2880s, 1955w, 1875w, 1810w, 1730w, 1605w, 1585w, 1495m, 1455s, 1370m, 1250–1200m (br.), 1140s, 1090s (br.), 1040s, 990s. $^1\text{H-NMR}$ (400 MHz): 7.33 (m, 15H); 4.67 (s, H–C(1)); 4.66 (d, A^1B^1 , $J = 12.5$, 1H, PhCH_2); 4.54 (d, A^1B^1 , $J = 12.5$, 1H, PhCH_2); 4.50 (d, A^2B^2 , $J = 12.0$, 1H, PhCH_2); 4.49 (d, A^3B^3 , $J = 12.0$, 1H, PhCH_2); 4.43 (d, A^2B^2 , $J = 12.0$, 1H, PhCH_2); 4.40 (d, A^3B^3 , $J = 12.0$, 1H, PhCH_2); 4.15 (dt, $J = 8.0, 4.0$, H–C(5)); 3.85 (dd, $J = 11.0, 4.0$, H–C(6)); 3.78 (m, H–C(3), H–C(4), H–C(6)); 3.68 (d, $J = 3.0$, H–C(2)); 3.37 (s, CH_3O). $^{13}\text{C-NMR}$ (50 MHz): 138.3 (s); 138.0 (s); 137.6 (s); 128.4–127.5 (6d); 100.2 (d); 75.3 (d); 72.7 (d); 72.4 (t); 72.3 (d); 71.9 (t); 71.3 (t); 67.4 (d); 62.7 (t); 55.3 (q). MS (CI): 463 ($M^+ - 1$), 431, 415, 397. Anal. calc. for $\text{C}_{28}\text{H}_{32}\text{O}_6$ (464.56): C 72.39, H 6.94; found: C 72.18, H 6.84.

2,3,4-Tri-O-benzyl-D-altrose (13). Prepared according to [19] (yield 11.1 g (86%), from 13.3 g of **12**). M.p. 85–86°, R_f (hexane/AcOEt 1:2) 0.29, $[\alpha]_D^{25} = -64.3^\circ$ ($c = 0.7$, CHCl_3). IR (KBr): 3540s, 3440s (br.), 3090w, 3060m, 3030m, 2970m, 2930m, 2880m, 1950w, 1815w, 1810w, 1605w, 1590w, 1495m, 1455s, 1380m, 1320m, 1280m, 1260m, 1220m, 1210m, 1080s, 1050s, 1000s, 950m, 900m, 860m, 780m, 750s, 730s, 700s. $^1\text{H-NMR}$ (200 MHz, $(\text{D}_6)\text{DMSO}$): 7.38–7.25 (m, 15H); 6.48 (d, $J = 7.0$, exchangeable with D_2O , OH–C(1)); 4.94 (d, $J = 7.0$, with D_2O : d, $J = 1.3$, H–C(1)); 4.70 (d, AB, $J = 12.2$, 1H, PhCH_2); 4.58 (s, PhCH_2); 4.53 (d, AB, $J = 12.2$, 1H, PhCH_2); 4.49 (d, AB, $J = 11.3$, 1H, PhCH_2); 4.42 (d, AB, $J = 11.3$, 1H, PhCH_2); 3.95 (dd, $J = 3.8, 2.7$, H–C(3)); 3.70–3.45 (m, 5H). $^{13}\text{C-NMR}$ (100 MHz, $(\text{D}_6)\text{DMSO}$): 138.8 (s); 138.6 (s); 138.4 (s); 128.1–127.3 (7d); 92.4 (d); 76.4 (d); 73.9 (d); 73.5 (d); 72.8 (d); 72.6 (t); 71.9 (t); 70.5 (t); 61.3 (t). Anal. calc. for $\text{C}_{27}\text{H}_{30}\text{O}_6$ (450.54): C 71.98, H 6.71; found: C 72.08, H 6.71.

2,3,4-Tri-O-benzyl-D-altrose-oxime (14). Compound **13** (8.64 g, 19.17 mmol) was added in one portion to a well-stirred mixture of $\text{NH}_2\text{OH} \cdot \text{HCl}$ (2.66 g, 38.33 mmol) and NaOMe (1.86 g, 43.50 mmol) in abs. MeOH (100 ml) and stirring was continued for 9 h at r.t. MeOH was then removed *i.v.* (bath temp. 40°), the residue was extracted with AcOEt ($5 \times$) and dried (MgSO_4). Evaporation of the solvent *i.v.* furnished 8.90 g of crude **14** as viscous sirup. R_f (hexane/AcOEt 1:2) 0.29.

N-(p-Nitrophenylmethylidene)2,3,4-tri-O-benzyl- β -D-altropyranosylamine N-Oxide (15). A mixture of crude **14** (8.90 g, 19 mmol), *p*-nitrobenzaldehyde (3.48 g, 23.0 mmol), TsOH (36 mg, 1 mol-%), and Drierite (40 g) in abs. CH_2Cl_2 (100 ml) was stirred at r.t. for 5 h. The mixture was then filtered through Celite and evaporated *i.v.* For analysis, a small amount of **15** was washed with 10% aq. NaHSO_3 soln. ($4 \times$) and H_2O ($3 \times$), dried (MgSO_4), and crystallised from hexane/AcOEt. M.p. 81–84°, R_f (hexane/AcOEt 1:2) 0.40, $[\alpha]_D^{25} = +112.4^\circ$ ($c = 0.57$, CHCl_3). UV: 253 (12122), 352 (18109). IR: 3605m, 3500m (br.), 3110w, 3090w, 3070m, 3030m, 3000m, 2930m, 2870m, 1955w, 1880w, 1810w, 1730w, 1600w, 1575m, 1520s, 1495m, 1455s, 1420m, 1340s, 1150s, 1100s, 900m, 865s, 840m. $^1\text{H-NMR}$ (200 MHz): 8.46, 8.27 (each d, $AA'BB'$, $J = 9.0$, C_6H_4); 7.91 (s, NO=CH); 7.39–7.03 (m, 3 PhCH_2); 5.38 (d, $J \leq 0.5$, H–C(1)); 4.58–3.86 (m, 12H); 2.95 (br. s, OH). $^{13}\text{C-NMR}$ (50.3 MHz): 147.8 (s); 137.4 (s); 137.2 (s); 135.4 (s); 130.9 (d); 129.5 (d); 128.4–127.7 (8d); 123.6 (d); 93.2 (d); 75.5 (d); 74.2 (d); 74.0 (t); 72.8 (t); 72.1 (d); 71.8 (d); 71.5 (t); 62.3 (t). MS (CI): 599 ($M^+ + 1$). Anal. calc. for $\text{C}_{34}\text{H}_{34}\text{N}_2\text{O}_8$ (598.66): C 68.21, H 5.72, N 4.68; found: C 68.20, H 5.65, N 4.61.

2,3,4-Tri-O-benzyl-1-deoxy-1-nitro-D-altropyranose (16) was prepared according to the procedure indicated for **8**. Yield after chromatography (toluene/AcOEt 8.5:1): 7.26 g (82%) of oily **16** as a mixture of anomers. R_f (toluene/AcOEt 8.5:1) 0.18 and 0.13, $[\alpha]_D^{25} = +74.3^\circ$ ($c = 0.53$, CHCl_3). IR: 3600m, 3450w (br.), 3090w, 3070w, 3030m, 3000m, 2930m, 2880m, 1955w, 1875w, 1815w, 1725w, 1605w, 1565s, 1495m, 1455m, 1370m, 1355m, 1330m, 1310m, 1240m, 1190m, 1165m, 1100s (br.), 1030s, 910m. $^1\text{H-NMR}$ (200 MHz): 7.39–7.11 (m, 15H); 5.65 (d, $J = 1.9$, H_α –C(1)); 5.37 (s, H_β –C(1)); $\alpha/\beta = 1:2$; 4.60–3.92 (m, 13H, all other H). $^{13}\text{C-NMR}$ (100 MHz): 101.1, 100.6 (2d, C(1) of both anomers); 76.4–70.8 (11 signals); 61.8 (t). MS (EI): 388 ($M^+ - 91$). Anal. calc. for $\text{C}_{27}\text{H}_{29}\text{NO}_7$ (479.53): C 67.63, H 6.09, N 2.92; found: C 67.40, H 6.20, N 2.80.

Dibenzyl 4,5,6-Tri-O-benzyl-1,2,3-trideoxy-3-nitro- α -D-altrio-3-octulopyranose-1-phosphonate (24). $\text{Bu}_4\text{NF} \cdot 3\text{H}_2\text{O}$ (1.19 g, 3.7 mmol) was added in one portion to a cooled (0°), stirred mixture of **16** (7.23 g, 15.07 mmol) and **18** (4.34 g, 15.07 mmol) in abs. THF (95 ml). Stirring was continued for 10 min at 0° , then for 2 h at r.t. Evaporation *i.v.* and chromatography (toluene/AcOEt 2:1) afforded 8.89 g (77%) of **24** (colorless oil, **24** decomposes partially on SiO_2). R_f (toluene/AcOEt 2:1) 0.17, $[\alpha]_D^{25} = +68.6^\circ$ ($c = 0.54$, CHCl_3). IR: 3600m, 3380m (br.), 3090m, 3060m, 3030m, 3000s, 2950m, 2920m, 2870m, 1955w, 1880w, 1810w, 1605w, 1585w, 1555s, 1495m, 1450s, 1370m, 1250–1200s (br.), 1160m, 1090s, 1050–990s (br.), 915m. $^1\text{H-NMR}$ (400 MHz): 7.45–7.10 (m, 25H); 5.02–4.86 (m, $(\text{PhCH}_2\text{O})_2\text{P}$); 4.47, 4.31 (A^1B^1 , $J = 11.8$, PhCH_2); 4.40, 4.23 (A^2B^2 , $J = 12.4$, PhCH_2); 4.39, 4.30 (A^3B^3 , $J = 11.7$, PhCH_2); 4.38 (d, $J = 3.6$, H–C(4)); 4.40 (m, H–C(7)); 3.94 (dd, $J = 12.5$, 2.5, H–C(8)); 3.88 (dd, $J = 10.3$, 2.5, H–C(6)); 3.79 (dd, $J = 12.5$, 3.3, H–C(8)); 3.59 (dd, $J = 3.6$, 2.5, H–C(5)); 2.48 (m, H–C(2)); 2.10 (m, H–C(2)); 1.90 (m, H–C(1)); 1.65 (br. s, OH); 1.55 (m, H–C(1)). $^{13}\text{C-NMR}$ (100 MHz): 137.7 (s); 137.2 (s); 136.5 (s); 136.2 (s); 128.7–127.8 (13d); 110.9 (d, $J(\text{C}, \text{P}) = 13.9$); 76.1 (d); 74.3 (d); 72.8 (d); 72.1 (t); 71.7 (t); 71.3 (t); 70.0 (d); 67.5 (dt, $J(\text{C}, \text{P}) = 6.2$); 67.30 (dt, $J(\text{C}, \text{P}) = 7.0$); 61.4 (t); 29.8 (dt, $J(\text{C}, \text{P}) = 2.4$); 19.4 (dt, $J(\text{C}, \text{P}) = 145.0$). $^{31}\text{P-NMR}$ (160 MHz): 32.2. Anal. calc. for $\text{C}_{43}\text{H}_{46}\text{NO}_{10}\text{P}$ (767.62): C 67.28, H 6.04, N 1.82, P 4.04; found: C 67.39, H 5.84, N 1.70, P 3.96.

Dibenzyl 4,5,6-Tri-O-benzyl-8-O-(dibenzylxyphosphoryl)-1,2,3-trideoxy-3-nitro- α -D-altrio-3-octulopyranose-1-phosphonate (25). Dibenzyl phosphorochloridate (9.47 g, 31.9 mmol) [**24**] was added dropwise under N_2 to a soln. of **24** (6.13 g, 7.98 mmol) in abs. pyridine (130 ml) at -30° . After 8 h at -10° , H_2O (5 ml) was added and stirring was continued for 15 min at 0° . Pyridine was removed *i.v.*, finally repeated by coevaporation with toluene. The residue was filtered through a short SiO_2 column (300 g) and then chromatographed (hexane/AcOEt 1:1) to afford 6.43 g (78%) of **25** as an oil. R_f (hexane/AcOEt 1:2) 0.39, $[\alpha]_D^{25} = +51.1^\circ$ ($c = 0.58$, CHCl_3). IR: 3090m, 3060m, 3030m, 3000s, 2950m, 2920m, 2890m, 2870m, 1955w, 1880w, 1810w, 1730w, 1605w, 1585w, 1555s, 1495m, 1450s, 1375m, 1340w, 1330w, 1305m, 1250s (br.), 1155m, 1095s, 1080s, 1000s (br.), 915m, 870m, 840m, 690m. $^1\text{H-NMR}$ (400 MHz): 7.34–7.08 (m, 35H), 4.99, 4.98 (d, $J(\text{C}, \text{P}) = 8.0$, $(\text{PhCH}_2\text{O})_2\text{P}$); 4.95–4.84 (m, $(\text{PhCH}_2\text{O})_2\text{PO}$); 4.49 (dm, $J = 10.3$, H–C(7)); 4.43, 4.25 (A^1B^1 , $J = 11.9$, PhCH_2); 4.36, 4.16 (A^2B^2 , $J = 12.2$, PhCH_2); 4.38 (m, H–C(8)); 4.36 (d, $J = 3.7$, H–C(4)); 4.28, 4.17 (A^3B^3 , $J = 11.7$, PhCH_2); 4.27 (m, H–C(8)); 3.82 (dd, $J = 10.3$, 2.4, H–C(6)); 3.54 (dd, $J = 3.7$, 2.4, H–C(5)); 2.45 (m, 1H); 2.15 (m, 2H); 1.40 (m, 1H). $^{13}\text{C-NMR}$ (100 MHz): 137.3–135.7 (5s); 128.6–127.7 (7d); 110.9 (d, $J(\text{C}, \text{P}) = 18.3$); 76.0 (d); 74.3 (t); 72.1 (t); 71.4 (t); 71.0 (d); 70.9 (d); 69.6 (d); 69.2 (t); 67.1 (t); 66.0 (t); 29.9 (t); 19.0 (dt, $J(\text{C}, \text{P}) = 143.2$). $^{31}\text{P-NMR}$ (160 MHz): +31.95 (phosphonate), –0.33 (phosphate). Anal. calc. for $\text{C}_{57}\text{H}_{59}\text{NO}_{13}\text{P}_2$ (1028.05): C 66.59, H 5.78, N 1.36, P 5.85; found: C 66.43, H 5.76, N 1.30, P 6.03.

Dibenzyl 4,5,6-Tri-O-benzyl-8-O-(dibenzylxyphosphoryl)-1,2-dideoxy-D-altrio-3-octulopyranose-1-phosphonate (26). A mixture of **25** (1.00 g, 0.97 mmol), HgCl_2 (2.65 g, 9.7 mmol), and H_2O (1.76 ml, 97 mmol) in CH_2Cl_2 (37 ml) was heated under reflux for 3 d. CH_2Cl_2 was evaporated *i.v.*, and the residue was taken up in CHCl_3 and filtered. Washing with 1% aq. KI soln. at 0° ($3\times$), then with H_2O ($3\times$), drying (MgSO_4), and evaporation *i.v.* furnished a yellowish oil, which, after chromatography (hexane/AcOEt 2:3), afforded 905.8 mg (95%) of **26** as a colorless oil. R_f (hexane/AcOEt 1:2) 0.15, $[\alpha]_D^{25} = +2.3^\circ$ ($c = 0.68$, CHCl_3). IR: 3440m (br.), 3090w, 3060m, 3030m, 2995s, 2950m, 2920m, 2890m, 2460w, 1950w, 1880w, 1810w, 1720w, 1605w, 1585w, 1495m, 1450s, 1375m, 1250–1200s (br.), 1155m, 1080s (br.), 1000s (br.), 915s. $^1\text{H-NMR}$ (400 MHz): 7.37–7.04 (m, 35H); 5.57 (s, OH); 5.01 (dd, $J = 7.7$, 1.1, $\text{PhCH}_2\text{OPO}_3$); 5.00 (d, $J = 7.8$, $\text{PhCH}_2\text{OPO}_3$); 4.92–4.87 (m, $(\text{PhCH}_2\text{O})\text{PO}$); 4.68, 4.45 (A^1B^1 , $J = 11.8$, PhCH_2); 4.45, 4.31 (A^2B^2 , $J = 11.7$, PhCH_2); 4.31 (m, 2H–C(8)); 4.27, 4.20 (A^3B^3 , $J = 11.9$, PhCH_2); 4.13 (dm, $J = 9.5$, H–C(7)); 3.80 (m, H–C(5), H–C(6)); 3.27 (d, $J = 3.2$, H–C(4)); 2.20, 1.70 (m, 2H–C(2), 2H–C(1)). $^{13}\text{C-NMR}$ (100 MHz): 137.7–136.1 (5s); 128.6–127.7 (10d); 97.8 (d, $J(\text{C}, \text{P}) = 18.6$); 76.5 (d); 74.1 (t); 73.8 (d); 73.0 (t); 72.2 (t); 71.6 (d); 69.2 (2t); 67.0–66.8 (4t); 29.9 (t); 19.1 (dt, $J(\text{C}, \text{P}) = 143.9$). $^{31}\text{P-NMR}$ (160 MHz): +35.0 (phosphonate), –0.5 (phosphate). Anal. calc. for $\text{C}_{57}\text{H}_{60}\text{O}_{12}\text{P}_2$ (999.05): C 68.53, H 6.05, P 6.20; found: C 67.99, H 6.43, P 6.00.

Tetrasodium 1,2-Dideoxy-8-O-phosphonato-D-altrio-3-octulose-1-phosphonate (27). A soln. of **26** (960 mg, 0.96 mmol) in 50 ml of dioxane/ H_2O 1:1 was treated with $\text{Pd}(\text{OH})_2/\text{C}$ (330 mg) [**25**] under 5 atm of H_2 for 3 h. After filtration and concentration of the filtrate to half of the volume, H_2O (25 ml) and fresh catalyst (300 mg) were added, and the hydrogenation was continued for 3 h. Filtration, treatment with *Dowex CCR-2* (Na^+ -form), lyophilisation and drying *i.v.* over P_2O_5 afforded 438 mg (100%) of **27**. R_f ($\text{PrOH}/\text{NH}_3/\text{H}_2\text{O}$ 4:3:1) 0.16, $[\alpha]_D^{25} = +3.6^\circ$ ($c = 0.83$, H_2O). $^1\text{H-NMR}$ (200 MHz, D_2O): 4.50–3.75 (m, 6H); 1.95 (m, 2H); 1.55 (m, 2H). $^{13}\text{C-NMR}$ (100 MHz, D_2O): β -D-furanose **27a**: 103.5 (d, $J(\text{C}, \text{P}) = 11.5$, C(3)); 80.9 (d, C(6)); 79.6 (d, C(4)); 75.6 (d, C(5)); 72.3 (d, C(7)); 65.0 (t, C(8)); 32.9 (dt, $J(\text{C}, \text{P}) = 3.7$, C(2)); 23.2 (dt, $J(\text{C}, \text{P}) = 129.8$, C(1)); α -D-furanose **27b**: 107.5 (d, $J(\text{C}, \text{P}) = 15.7$, C(3)); 83.7 (d, C(4)); 81.4 (d, C(6)); 77.4 (d, C(5)); 71.2 (d, C(7)); 65.3 (t, C(8)); 29.6 (t, C(2)); 23.2 (dt, $J(\text{C}, \text{P}) = 129.8$, C(1)); α -D-pyranose **27c**: 100.5 (d, C(3)); 72.1 (d, C(5)); 69.4 (d, C(7)); 68.8 (d,

C(4)); 64.0 (*d*, C(6)); 63.9 (*t*, C(8)); 32.1 (*t*, C(2)); 23.2 (*dt*, J(C, P) = 129.8, C(1)). ³¹P-NMR (160 MHz, D₂O): 22.8 (27b), 22.3 (27a), 21.3 (27c); integral over phosphonate signals 27b/27a/27c = 35.9:58.2:5.9; 5.5 (27b), 5.2 (27a), 4.7 (27c). MS (FAB): 457 (*M* + 1), 435 (*M* + 2 - Na), 413 (*M* + 3 - 2Na), 391 (*M* + 4 - 3Na), 369 (*M* + 5 - 4Na). Anal. calc. for C₈H₁₄Na₂O₁₂P₂ (456.10): C 21.07, H 3.09, P 13.58; found: C 20.88, H 3.38, P 13.39.

REFERENCES

- [1] B. Aebischer, R. Meuwly, A. Vasella, *Helv. Chim. Acta* **1984**, *67*, 2236.
- [2] R. Engel, *Chem. Rev.* **1977**, *77*, 349.
- [3] K. H. Scheit, in 'Nucleotide Analogues: Synthesis and Biological Function', John Wiley and Sons, New York, 1980, Chap. 4.
- [4] J. C. Tang, B. E. Tropp, R. Engel, *Tetrahedron Lett.* **1978**, 723.
- [5] F. M. Unger, D. Stix, E. Möderndorfer, F. Hammerschmid, *Carbohydr. Res.* **1978**, *67*, 349.
- [6] G. H. Jones, J. G. Moffatt, **1971**, U. S. Patent 3,583,974.
- [7] R. W. Mclard, *Tetrahedron Lett.* **1982**, 2631.
- [8] F. Nicotra, F. Ronchetti, G. Russo, *J. Org. Chem.* **1982**, *47*, 4459.
- [9] Yu. A. Zhdanov, Z. I. Glebova, L. A. Uzlova, N. I. Vikrishchuk, *Zh. Obshch. Khim.* **1983**, *53*, 2616.
- [10] M. Chmielewski, J. N. Bemiller, D. P. Cerretti, *Carbohydr. Res.* **1981**, *97*, C 1.
- [11] B. Aebischer, Diss No. 584, Univ. Fribourg (Switzerland), 1983.
- [12] B. Aebischer, A. Vasella, *Helv. Chim. Acta* **1983**, *66*, 789.
- [13] R. Julina, Diploma work, Univ. Zürich (Switzerland), 1982.
- [14] S. Mirza, A. Vasella, *Helv. Chim. Acta* **1984**, *67*, 1562.
- [15] R. Barker, D. L. MacDonald, *J. Am. Chem. Soc.* **1960**, *82*, 2301.
- [16] John W. Van Cleve, *Carbohydr. Res.* **1982**, *106*, 239.
- [17] P. P. Singh, M. M. Gharia, F. Dasgupta, H. C. Srivastava, *Tetrahedron Lett.* **1977**, 439.
- [18] C. P. J. Glaudemans, H. G. Fletcher jr., in 'Synthetic Procedures in Nucleic Acid Chemistry', Ed. W. W. Zorbach and R. S. Tipson, Interscience Publish., 1968, Vol. 1, p. 126.
- [19] N. K. Richtmyer, in 'Methods in Carbohydrate Chemistry', Ed. R. L. Whistler and M. L. Wolfrom, Academic Press, New York, 1962, Vol. 1, p. 107.
- [20] K. Sasse, in 'Houben-Weyl: Methoden der organischen Chemie', Ed. E. Müller, Thieme, Stuttgart, 1963, Vol. XII/1, p. 401.
- [21] a) N. Minova, M. Hirayama, S. Fukatsu, *Tetrahedron Lett.* **1984**, 1147; b) 'Phosphorus Chemistry', Proceedings of the 1981 International Conference, Ed. L. D. Quin and J. G. Verkade, p. 243.
- [22] S. J. Angyal, G. S. Bethell, *Aust. J. Chem.* **1976**, *29*, 1249.
- [23] T. A. W. Koerner, L. W. Cary, N. S. Bhacca, E. S. Younathan, *Biochem. Biophys. Res. Commun.* **1973**, *51*, 543.
- [24] F. R. Atherton, *Biochem. Prep.* **1957**, *5*, 1.
- [25] W. M. Pearlman, *Tetrahedron Lett.* **1967**, 1663.
- [26] F. P. Franke, M. Kapuscinski, J. K. McLeod, J. F. Williams, *Carbohydr. Res.* **1984**, *125*, 177.
- [27] B. Rayner, C. Tapiero, J.-L. Imbach, *Carbohydr. Res.* **1976**, *47*, 195.
- [28] M. Mac Coss, M. J. Robins, B. Rayner, J.-L. Imbach, *Carbohydr. Res.* **1977**, *59*, 575.
- [29] R. U. Lemieux, J. D. Stevens, *Can. J. Chem.* **1966**, *44*, 249.
- [30] C. Altona, C. A. G. Haasnoot, *Org. Magn. Reson.* **1980**, *13*, 417.
- [31] F. Baumberger, A. Vasella, *Helv. Chim. Acta* **1983**, *66*, 2210.
- [32] B. Aebischer, A. Vasella, H. P. Weber, *Helv. Chim. Acta* **1982**, *65*, 621.
- [33] M. Martin-Lomas, M. E. Chacon-Fuertes, *Carbohydr. Res.* **1977**, *59*, 604.
- [34] R. J. M. Nolte, J. A. J. van Zomeren, J. W. Zwikker, *J. Org. Chem.* **1978**, *43*, 1972.
- [35] P. Boullanger, D. Marmet, G. Descotes, *Tetrahedron* **1979**, *35*, 163.
- [36] H. Steinlin, L. Camarda, A. Vasella, *Helv. Chim. Acta* **1979**, *62*, 378.
- [37] B. M. Aebischer, H. W. Hanssen, A. T. Vasella, W. B. Schweizer, *J. Chem. Soc., Perkin Trans. 1* **1982**, 2139.
- [38] N. Cohen, B. L. Banner, R. J. Lopresti, F. Wong, M. Rosenberger, Y. Liu, E. Thom, A. A. Liebmann, *J. Am. Chem. Soc.* **1983**, *105*, 3661.
- [39] V. Kohli, H. Bloecker, H. Koester, *Tetrahedron Lett.* **1980**, 2683.